

Research Article

Isolation of Thiamine-Binding Protein from Black Glutinous Rice Bran (*Oryza sativa var. Glutinosa***) Using Ammonium Sulfate Precipitation and Equilibrium Dialysis**

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Abstract: Thiamine is an essential cofactor in energy metabolism. Deficiency of this vitamin can cause disorders in the nervous and cardiovascular systems. Conventional methods for examining thiamine levels in the body tend to be expensive. One solution is to use a technique similar to ELISA (Enzyme-Linked Immunosorbent Assay), utilizing a specific thiamine-binding protein as a substitute for antibodies. This study aims to investigate the presence of thiamine-binding protein in black glutinous rice bran (Oryza sativa var. Glutinosa). The methods used include salting out, dialysis, and equilibrium dialysis. The results show that the thiamine-binding protein (TBP) in black glutinous rice bran precipitated with 90% ammonium sulfate. The average total protein was 5.732 mg/mL after dialysis, and equilibrium dialysis demonstrated that TBP can bind thiamine with a binding capacity ranging from 30 to 76.6 µg per 10 grams of sample.

Keywords: Black Glutinous Rice Bran, Equilibrium Dialysis, Isolation, Thiamine-Binding Protein

Introduction

Thiamine is a vitamin B group that cannot be synthesized by the human body and must be obtained from dietary sources μ . Thiamine plays a crucial role in carbohydrate metabolism and energy production, primarily as a coenzyme in the oxidative decarboxylation of pyruvate and the citric acid cycle. Thiamine is also essential for normal nerve function as it is involved in neurotransmitter production and myelin maintenance $[2]$. The requirement for this vitamin varies among individuals, influenced by age, energy needs, carbohydrate intake, and body mass. Based on these considerations, the Food and Nutrition Board of the US Institute of Medicine recommends a thiamine intake of 0.5 mg/1000 kcal $[3]$.

 Thiamine deficiency impedes or severely restricts thiamine-dependent reactions, resulting in substrate accumulation for these reactions. This condition leads to decreased activity of the pyruvate dehydrogenase and α -ketoglutarate dehydrogenase enzymes and impaired adenosine triphosphate (ATP) synthesis. Reduced ATP synthesis in critical areas such as the nervous and cardiovascular systems can lead to cell death, Wernicke's encephalopathy, and impaired insulin synthesis and secretion \mathbb{H} . Although thiamine deficiency is rare in various countries worldwide, due to its limited storage in the body and short half-life, a sufficient and regular supply from the diet is necessary. Under conditions of starvation or malnutrition, thiamine deficiency can trigger the onset of beriberi as the first disease to appear [\[4\]](#page-8-3). Therefore, it is crucial to examine thiamine levels in the human body.

 Methods that accurately detect thiamine levels in biological fluids are needed to screen for thiamine deficiency. Several techniques can be used to detect thiamine levels in serum, including spectrophotometry, high-performance liquid chromatography (HPLC), and enzyme-linked immunosorbent assay (ELISA) [\[5,](#page-0-0)[6,](#page-8-4)[7\]](#page-8-5). Some existing methods require special equipment and are costly to implement, hence the development of a serum thiamine assay method using a principle similar to ELISA, which utilizes thiamine-binding protein as an antibody substitute.

 Several studies report that thiamine-binding protein (TBP) is found in various materials, including bacteria and other microorganisms $^{[8,9,10,11]}$ $^{[8,9,10,11]}$ $^{[8,9,10,11]}$ $^{[8,9,10,11]}$ $^{[8,9,10,11]}$ $^{[8,9,10,11]}$. TBP has also been isolated from egg white, yolk, and animal tissues $[12,13]$ $[12,13]$. Thiamine-binding protein can be obtained from plants that are readily available locally. Some

sources of thiamine-binding protein that have been studied include mung beans, corn bran, and sunflower seeds $[8,14,15,16]$ $[8,14,15,16]$ $[8,14,15,16]$ $[8,14,15,16]$. This study aims to isolate and identify the presence of thiamine-binding protein in black glutinous rice bran (*Oryza sativa var. Glutinosa*).

 Methods that accurately detect thiamine levels in biological fluids are needed to screen for thiamine deficiency. Several techniques can be used to detect thiamine levels in serum, including spectrophotometry, high-performance liquid chromatography (HPLC), and enzyme-linked immunosorbent assay (ELISA) [5.6.[7\]](#page-8-5). Existing methods often require specialized equipment and are expensive to implement. Therefore, developing a serum thiamine testing method using a principle similar to ELISA, which utilizes a specific thiamine-binding protein as a substitute for antibodies, presents a potential solution.

 Several studies report that thiamine-binding proteins (TBP) are found in various materials, including bacteria and other microorganisms [8-11]. TBP has also been isolated from egg white, yolk, and animal tissues $[12,13]$ $[12,13]$. Thiamine-binding protein can be sourced from plants that are readily available in Indonesia. Some sources of TBP that have been studied include mung beans, corn bran, and sunflower seeds $[8,14,15,16]$ $[8,14,15,16]$ $[8,14,15,16]$ $[8,14,15,16]$.

 This study aims to isolate and identify the presence of thiamine-binding protein in black glutinous rice bran (*Oryza sativa var. Glutinosa*). Rice bran is a byproduct of rice milling. The bran consists of the rice husk, which is typically discarded during the rice polishing process. Although rice bran makes up only 1%–2% of the entire grain, it contains 70% of the grain's total nutrition $[21]$. Currently, rice bran is more commonly used as animal feed. In this study, its utilization as a source of thiamine-binding protein offers a cheaper alternative. Plant-based thiamine-binding proteins hold great potential due to their abundant availability, easy accessibility, safety, sustainability, and simpler, more cost-effective isolation process compared to animal or microbial sources.

Materials and Methods

Materials

 The black glutinous rice bran used in this study was obtained from the traditional Rawamangun market and is commonly consumed by Indonesians. Other materials used include Bovine Serum Albumin (BSA) as a protein standard (Nacalai Tesque), Na₂HPO₄ (Merck), KH₂PO₄ (Merck), BaCl₂ (Merck), NaCl (Merck), (NH4)2SO⁴ (Merck), and distilled water. The equipment used includes a balance with a 220-gram capacity (Sartorius), a blender, a 150 R sieve, a centrifuge (Hettich), a pH meter (Schott), a magnetic stirrer (Thermolyne), a UV-Vis spectrophotometer (Thermo Fisher), 1 mL and 10 µL micropipettes (Bio-Rad), 1 mL and 10 µL microtips (Extragene), 25 mL microtubes (Extragene), Falcon tubes (Extragene), 33 mm wide cellophane bags (Sigma-Aldrich), and several glassware such as test tubes and beakers (Pyrex).

Methods

Sample and Buffer Preparation

 One hundred grams of black glutinous rice bran were ground using a blender until finely powdered and then sieved using a 150 R sieve. A 1-liter 0.05 M phosphate buffer with pH 7 was prepared by dissolving 0.53 grams of KH₂PO₄, 1.09 grams of Na₂HPO₄, and 4.5 grams of NaCl in distilled water while stirring with a magnetic stirrer. The pH was adjusted to 7 by adding NaOH or HCl solution.

Protein Extraction (Salting Out)

 Thiamine-Binding Protein (TBP) was extracted from black glutinous rice bran samples, with three repetitions (repetition 1, 2, and 3) carried out. Each repetition weighed 10 grams of black glutinous rice bran powder, which was added to 100 mL of 0.05 M phosphate buffer solution at pH 7. The mixture was stirred using a magnetic stirrer until homogeneous for 24 hours at 4°C. To minimize protein degradation, the entire protein isolation process was conducted at 4°C. The homogenates from the three repetitions were centrifuged at 28,000 g for 15 minutes, and the supernatant (crude extract) was collected for the next stage. The protein content of each crude extract was measured using a spectrophotometer, and its concentration was calculated using a standard protein curve.

 The protein in each supernatant sample was precipitated using the salting out method by adding ammonium sulfate powder until 90% saturation was reached $[5]$. Ammoniumsulfate is highly effective in maintaining protein stability during the purification process, thus reducing protein denaturation during the extraction stage. Ammonium sulfate powder was added gradually while stirring with a magnetic stirrer, then centrifuged at 28,000 g for 15 minutes $[5]$. The precipitate obtained was redissolved to the original volume by adding phosphate buffer.

Protein Assay

Each isolation step was tested for protein using the Warburg-Christian method (λ 280 nm) with bovine serum albumin (BSA) as the standard $[17]$.

Standard Protein Curve

 The standard protein curve was made by preparing a series of BSA concentrations. The concentrations used were 0 μ g/mL, 50 μ g/mL, 100 μ g/mL, $\overline{200 \mu}$ g/mL, and so on up to 1000 μ g/mL. Each concentration was prepared in duplicate, and the average absorbance at 280 nm was measured and plotted.

Standard Thiamine Curve

 The standard thiamine curve was made by dissolving 500 mg of thiamine powder in distilled water to a volume of 50 mL as stock. A series of concentrations were prepared from the stock solution by dilution, starting from 0 μ g/mL, 2 μ g/mL, 4 μ g/mL, 6 μ g/mL, and so on up to 20 μ g/mL. Each concentration was prepared in duplicate, and the average absorbance at 235 nm was measured and plotted. **Dialysis**

 Each fraction from the salting out was dialyzed using cellophane membrane bags with a 0.05 M phosphate buffer exchange at pH 7 to separate the protein from the remaining ammonium sulfate. Dialysis was considered complete when the ammonium sulfate concentration in the dialysis buffer outside the bag was zero, using a sulfate test (1 mL solution outside the bag + 1 mL HCl + 1 mL BaCl₂).

Equilibrium Dialysis (TBP Binding Capacity Test)

 The thiamine-binding capacity of TBP was tested using the equilibrium dialysis method. Each salting out fraction after dialysis was sampled at 198 mL to be placed in cellophane membrane bags, adding 0.02 mL of 10 mg/mL thiamine solution, and mixed until homogeneous. The bags were then placed in beakers containing 100 mL of distilled water. All bags had to be submerged. The buffer outside the bag was measured for thiamine concentration at 10-minute intervals at 235 nm, stopping the reading once equilibrium was reached, indicated by constant absorbance values.

Results

 Black glutinous rice is very popular among Indonesians and is usually processed into snacks such as black glutinous rice bran porridge or black glutinous rice tape. Its relatively complete nutritional content makes it a good source of calories and vitamins. Table 1 shows the nutritional content of black glutinous rice bran (100 grams). Table 1: Nutritional Content of Black Glutinous Rice (100 grams)

Compiled by author

Protein Standard Curve

 The protein standard curve was prepared by preparing a series of BSA concentrations and measuring protein levels using the Christian Welburg method at a wavelength of 280 nm. The protein standard curve will be used to measure protein concentrations. The standard curve equation is $y=0.0007x+0.004$; with a correlation coefficient $R^2 = 0.9999$. The standard curve graph can be seen in Figure 1.

Figure 1. BSA Standard Curve

Thiamine Standard Curve

 The thiamine standard curve was created by preparing a series of thiamine concentrations ranging from 0 to 10 µg/mL, read at the maximum wavelength of thiamine, 235 nm. The linear equation of the thiamine standard curve is y=0.0427x+0.0094; with a correlation coefficient R^2 =0.9969. The thiamine standard curve will be used to measure thiamine concentrations during equilibrium dialysis. The standard curve graph is shown in Figure 2.

Figure 2. Thiamine Standard Curve

Protein Precipitation

 The protein content of the three crude fractions from homogenized black glutinous rice samples was measured at 280 nm and calculated using the BSA standard curve. Table 2 shows the protein content of the crude fractions from each sample.

 The protein content of the three crude fractions from the homogenized black glutinous rice bran sample repetitions was measured at a wavelength of 280 nm and calculated using the BSA standard curve. Table 2 shows the protein content of the crude fractions for each sample repetition. It is evident that each sample

repetition has varying protein levels, as the protein content in each 10 grams of black glutinous rice bran powder taken also differs.

Salting Out and Dialysis

 The protein from the crude extract was precipitated using the salting out method with ammonium sulfate at 90% saturation, then dialyzed to remove any remaining salts. Table 3 shows the protein content of each sample after salting out and dialysis.

Salting Out Sample	Absorbance $(\lambda = 280 \text{nm})$	Protein Concentration 100x Dilution $(\mu g/mL)$	Protein Concentration $(\mu g/mL)$	Protein Concentration Percentage $(\%) / 10$ gram
Repetition 1	0,037	47,14	4714,29	0,047
Repetition 2	0,040	51,43	5142,86	0,051
Repetition 3	0,044	57,14	5714,29	0,057

Table 3. Total Protein of Salting Out Fractions After Dialysis

Equilibrium Dialysis

 Equilibrium dialysis was conducted on the three samples to determine the presence of thiamine-binding protein and the concentration of thiamine it could bind. Thiamine absorbance was read in the solution outside the cellophane bag at 235 nm during equilibrium dialysis. Repetition 1 reached equilibrium at 120 minutes with an absorbance reading of 0.029 (Table 4). Repetition 2 reached equilibrium at 140 minutes with an absorbance reading of 0.069 (Table 5). Repetition 3 reached equilibrium at 140 minutes with an absorbance reading of 0.062 (Table 6). The three samples, after reaching equilibrium, are shown as a flat line in Figures 3, 4, and 5.

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Figure 3. Equilibrium Dialysis Graph for Repetition 1. Equilibrium reached at the 120th minute with an absorbance reading of 0.029 that is marked with blue (Table 4)

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Time	Absorbance	Time	Absorbance			
(minutes)	$(\lambda=280 \text{ nm})$	(minutes)	$(\lambda = 280 \text{ nm})$			
0						
10	0,019	90	0,056			
20	0,027	100	0,058			
30	0,033	110	0,061			
40	0,037	120	0,066			
50	0,044	130	0,067			
60	0,045	140	0,069			
70	0,049	150	0,069			
80	0,052	160	0,069			

Figure 4. Equilibrium Dialysis Graph for Repetition 2. Equilibrium reached at the 140th minute with an absorbance reading of 0.069 that is marked with blue (Table 5).

Time (minutes)	Absorbance $(\lambda=280 \text{ nm})$	Time (minutes)	Absorbance $(\lambda=280 \text{ nm})$	
0	0			
10	0,02	90	0,047	
20	0,021	100	0,051	
30	0,028	110	0,054	
40	0,039	120	0,057	
50	0,036	130	0,057	
60	0,043	140	0,062	
70	0,042	150	0,062	
80	0,048	160	0,062	

Table 6. Equilibrium Dialysis Repetition 3

Figure 5. Equilibrium Dialysis Graph for Repetition 3. Equilibrium reached at the 140th minute with an absorbance reading of 0.062 that is marked with blue (Table 6).

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 Absorbance readings at the end of equilibrium dialysis for the three samples were recorded and converted to thiamine concentrations using the thiamine standard curve. The thiamine concentration was then calculated using the formula (1) to determine the thiamine concentration bound to the thiamine-binding protein in the cellophane bag. The calculation results can be seen in Table 7

> (1) [Bound Ligand] = $\frac{Total$ ligand Mol−Free ligand Mol The volume of the compartment in the cellophane bag

 Total ligand moles are the amount of thiamine moles introduced into the cellophane bag. Free ligand moles are obtained from the thiamine concentration in the outer compartment multiplied by the total volume of both compartments. The total ligand moles used in this study are: 0,2

Discussion

 Several studies have isolated thiamine-binding proteins from other sources. One such study by Gunarti et al. isolated thiamine-binding protein from mung beans, achieving a total protein concentration of 2.5 mg/ml after dialysis $[5]$. When compared to mung beans, black glutinous rice bran, with an average total protein concentration of 5.732 mg/mL after dialysis, has twice the concentration, suggesting a higher potential for isolating thiamine-binding protein from black glutinous rice bran.

 Another source of thiamine-binding protein studied is corn. The total protein obtained from corn in the crude sample was 792 mg $^{[15]}$ $^{[15]}$ $^{[15]}$. In comparison, the average total protein from black glutinous rice bran in the crude sample was 750 mg, which is slightly less. However, another study by Mitsunaga et al. found that the thiamine-binding capacity of corn protein was not as good as black glutinous rice bran, with 20.5 µg per 10 grams of sample compared to black glutinous rice, which ranged from 30-76.6 µg per 10 grams of sample [19] .

 Thiamine-binding protein has also been isolated from buckwheat (*Fagopyrum esculentum*), with a total protein of 292 mg after salting out and dialysis ^{[\[20\]](#page-8-14)}. In comparison, black glutinous rice provided twice as much total protein at the salting out stage, with 593 mg.

There are many other sources of thiamine-binding protein. Mitsunaga et al. $[19]$ studied 18 plant species containing thiamine-binding protein. Plants with thiamine-binding capacities comparable to black glutinous rice bran (30-76.6 µg per 10 grams of sample) include soybeans (34.8 µg), brown rice (44.5 µg), *Vigna sinensis L*. (46.4 µg), peas (52.5 µg), *Phaseolus vulgaris L. var. cultiva Kintoki* (57.9 µg), *Phaseolus vulgaris L. var. cultiva Aschers* (61.9 µg), and *Phaseolus coccineus L. var. albus Bailey* (64.5 µg) [\[19\]](#page-8-14) . Plants exceeding the binding capacity of black glutinous rice bran (>76.6 µg per 10 grams of sample) include wheat germ (87.6 µg), *Fagopyrum esculentum* Moench. (104.1 µg), bran (147.7 µg), sunflower seeds (288.5 µg), and sesame seeds (675.0 µg) ^{[\[19\]](#page-8-14)}.

 The thiamine-binding protein isolated from black glutinous rice bran in this study is still impure. Further purification steps are needed to obtain pure TBP. Pure TBP can be used as a substitute for antibodies in a thiamine measurement tool similar to ELISA. This tool can be used to measure thiamine levels in food and biological fluids.

Conclusion

 Thiamine-binding protein can be found in extracts of black glutinous rice bran and has been shown to bind thiamine effectively.

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References

- [1] Page GLJ, Laight D, Cummings MH. Thiamine deficiency in diabetes mellitus and the impact of thiamine replacement on glucose metabolism and vascular disease. Int J Clin Pract. 2011 Jun;65(6):684–90.
- [2] Sullivan, K. (2002). Vitamins and minerals: A practical approach to a health diet and safe supplementation.
- [3] *Food and Nutrition Board US Institute of Medicine* (The National Academies Press. Nutrition - Dietary Reference Intakes: DRIs): Recommended Dietary Allowances and Adequate Intakes, Vitamins. Available at
- [4] Román GC, Román GC. Ariza Prison: Cienfuegos, Cuba, August 5, 1992. Cuba Blind. 2016 Jan 1;11–8.
- [5] Gunarti DR, Rahmi H, Sadikin M. Isolation and Purification of Thiamine Binding Protein from Mung Bean. HAYATI J Biosci. 2013 Mar 1;20(1):1–6.
- [6] Anyakora C, Afolami I, Ehianeta T, Onwumere F. HPLC analysis of nicotinamide, pyridoxine, riboflavin and thiamin in some selected food products in Nigeria. Afr J Pharm Pharmacol; 2008:2(2):29–36.
- [7] Chen Y, Tian F. Enzymatic Catalytic Spectrophotometric Determination of Thiamine in Food. Food Anal Methods. 2010 Mar 28;3(1):7–11.
- [8] Wingfield P. Protein precipitation using ammonium sulfate. Curr Protoc protein Sci. 2001 May;Appendix 3:Appendix 3F.
- [9] Rosenberg IM. Protein analysis and purification. 2nd ed. Boston: Birkhäuser; 1996. Chapter 5, Getting started with protein purification; p. 118–52.
- [10] Simonian MH. Spectrophotometric Determination of Protein Concentration. Curr Protoc Cell Biol. 2002 Jul;15(1):A.3B.1-A.3B.7.
- [11]Barwick V. Preparation of calibration curves: a guide to best practice. LGC/VAM/2003/032. Available from: https://www.lgcgroup.com/media/1735/prepration-of-calibration-curves_aguide-to-best-practice.pdf.
- [12]Analytical chemistry: calibration curves. Cambridge, MA: JoVE Science Education Database; 2019 [cited 2091 Oct 28]. Available from: https://www.jove.com/scienceeducation/10188/calibration-curves.
- [13] N K. Methods in enzymology. Acad Press. 1968;I:76.
- [14] Shimizu M, Yoshida T, Toda T, Iwashima A, Mitsunaga T. Isolation of a Thiamine-binding Protein from Rice Germ and Distribution of Similar Proteins. Biosci Biotechnol Biochem. 1996 Jan 12;60(3):453–7.
- [15]Adachi T, Watanabe K, Mitsunaga T. Characterization of thiamin-binding protein from maize seeds. Plant Physiol Biochem. 2001 Feb 1;39(2):99–105.
- [16]Watanabe K, Konishi A, Mitsunaga T. Molecular characteristics of thiamin-binding protein from sunflower seeds. Plant Physiol Biochem. 2002 May 1;40(5):417–21
- [17] Rosenberg IM. 2005. Protein Analysis and Purification. Boston: BirkhÓuser.
- [18]Mohammad F. Hossain, Mamoon Rashid, Rajjit Sidhu, Randy Mullins, and Susan L. A Simplified, Specific HPLC Method of Assaying Thiamine and Riboflavin in Mushrooms., Int J Food Sci. 2019; 2019: 8716986. Published online 2019 Feb 3. doi: 10.1155/2019/8716986, PMCID: PMC6378034, PMID: 30854396
- [19]Mitsunaga T, Shimizu M, Iwashima A. Occurrence of thiamine-binding proteins in plant seeds. J Plant Physiol. 1986 Jun 1;124(1–2):177–80.
- [20] Mitsunaga T, Matsuda M, Shimizu M, Iwashima A. Isolation and properties of a thiaminebinding protein from buckwheat seed. Cereal Chem. 1986;63:332–5.
- [21]Fuller, D. Q., & Harvey, E. L. (2006). The archaeobotany of Indian pulses: identification, processing and evidence for cultivation. *Environmental Archaeology*, *11*(2), 219-246

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