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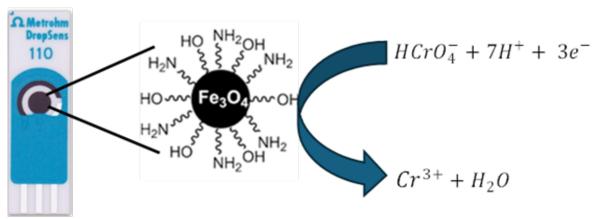
Modification of Screen-Printed Carbon Electrode (SPCE) by Magnetic Fe₃O₄-Chitosan for Detection of Hexavalent Chromium

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GRAPHICAL ABSTRACT



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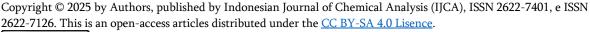
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ABSTRACT

A new modification of screen-printed carbon electrode (SPCE) with magnetic Fe₃O₄-chitosan has been developed. Magnetic Fe₃O₄-chitosan was made from a mixture of Fe²⁺/Fe³⁺ (1:2) with 1% chitosan. Sodium tripolyphosphate was used as a crosslinking reagent for chitosan under alkaline conditions using NH₃. SPCE modification with Fe₃O₄ can increase the sensitivity of Cr(VI) detection by differential pulse voltammetry (DPV). This method resulted in a sensitivity of 21.9 ppm/ μ A in the 0.1 - 1 ppm Cr(VI) concentration range in a 0.005 M KCl-HCl electrolyte mixture. The detection limit (LoD) is 0.2 ppm with an average accuracy of 94%.

1. INTRODUCTION

Chromium ion has both positive and negative effects on human health depending on absorption, exposure time, and degree of oxidation. The Cr(III) form is an essential nutrient for humans, and according to the World Health Organization [1], the ideal daily intake is between 50 and 200 g per day for carbohydrate, protein, and fatty acid metabolism. However, its excess in the body causes serious health problems. In contrast, Cr(VI) ion is 10-100 times more dangerous than Cr(III) because when it is accumulated in the human body, it leads to cancer, liver, and kidney damage. The Cr(VI) may also cause skin irritation and be considerably toxic to the protoplasm of living things [2].





Determination of chromium content is generally expressed in terms of total chromium, although chromium ions have different toxicity levels. Several methods for determining the concentration of chromium are atomic absorption spectrometry (AAS), atomic emission spectrometry (AES), X-ray spectroscopy, inductively coupled plasma mass spectrometry (ICP-MS), high-pressure liquid chromatography (HPLC) [3–7]. The determination of chromium as Cr(VI) can be done using a diphenyl carbazide reagent. However, oxidation must be carried out to determine Cr(III) [8–10]. Meanwhile, the electrochemical method of Cr(VI) can be detected specifically and, in potentiometry, has a higher detection limit than the threshold value [11–14]. Voltammetrically, Cr(VI) can be reduced to Cr(III) in acidic conditions, so it is not determined as total chromium. Therefore, the proposed method for determining hexavalent chromium ions in this study uses an electrochemical voltammetric sensor.

Hexavalent chromium electrochemical sensors have been developed using the voltammetric stripping technique with various working electrodes, either with or without modification of other materials [15–21]. Several voltammetric methods of determining both Cr(III) and Cr(VI) species have been developed, and stripping voltammetry is generally used. The Au-Ag-Pt electrode was used as the working electrode for the determination of Cr(VI) by differential pulse cathodic stripping voltammetry (DPCSV) [15]. An electrochemical sensor for determining Cr(VI) using SPE surface-modified gold nanoparticles (AuNPs). SPE modification by casting pure AuNPs increased the sensitivity to detect Cr(VI) ions using anodic stripping voltammetry. The Cr(VI) ion was reduced to metal chromium on SPE-AuNPs by applying a precipitation potential of -1.1 Volt for 180 seconds. After that, the peak chromium oxidation current was obtained by linear sweep voltammetry in the range of -1.0 to 0.2 Volt [16, 17]. Moreover, AuNPs/rGO Nanocomposite [18], a nanocomposite of gold nanoparticles deposited on the sidewall of a carbon nanotube, Ox-MWCNT-Au, was reported and prepared using simple chemical reduction. Carbon nanotube-gold was used by cyclic voltammetry, and Cr(VI) reduction was observed at a peak potential of 0.52 V compared to SCE in acidified H₂SO₄ solution, pH 2.0 [19]. Iron-based materials have increased the sensitivity of detecting hexavalent chromium ions, including nickel hexacyanoferrate, as a modifier for glassy carbon electrodes [20]. Cr³⁺ ions can be reduced to Cr²⁺ at a potential of -0.408 volts, and Cr₂O₇²⁻ ions are reduced to Cr³⁺ at a potential of +1.33 volts under acidic conditions. Determination of hexavalent chromium by voltammetry has also been carried out with a modified glassy carbon electrode of Fe3O4starch composite as the working electrode. The Cr(VI) is reduced to Cr(III) at a potential of 0.99 volts under acidic conditions with a supporting electrolyte of 0.05 M H₂SO₄-K₂SO₄. The electrode detection limit is 3 ppm, less sensitive, and disposable [21]. The two reduction potentials are so far apart that trivalent and hexavalent chromium ions can be determined simultaneously [22].

Therefore, in this study, a modification of the screen-printed carbon electrode with Fe3O4-chitosan composite was carried out. Magnetite (Fe₃O₄) is a superparamagnetic metal oxide that can increase electron transfer. Modifying carbon electrodes with magnetite has been studied to determine paracetamol voltammetrically [23]. Meanwhile, chitosan can adsorb Cr(VI), with an adsorption capacity of 212.8 mg/g chitosan [24]. Chitosan is a natural polymer that can form nanoparticles with sodium tripolyphosphate (Na-TTP). Chitosan dissolved in acid was reacted with Na-TTP in an alkaline solution [25, 26]. Chitosan nanoparticles were made within 2 hours using the ionic gelation method. Chitosan molecules in the form of gel contact with poly-anions due to the formation of inter- and intramolecular cross-links mediated by poly anions. Chitosan nanoparticles were made by adding a negatively charged Na-TPP solution with a positively charged chitosan solution while stirring at room temperature. The ratio of chitosan to Na-TTP will determine the size of the resulting nanoparticles [27].

2. EXPERIMENTAL

2.1 Reagents and apparatus

The reagents used were of high purity and used as received. Double-distilled water was used for all tests. Chemicals used include: KCl (Merck), HCl (Merck, 32%; 1.18g/mL), NH₄OH (Merck, 25%; 0.9 g/mL), K₂Cr₂O₇, FeSO₄.7H₂O, FeCl₃.6H₂O, chitosan powder (Merck, DAC Degree \geq 80%), CH₃COOH (Merck, 99.5–100%; 1.05 g/mL), Na-tripolyphosphate (Merck). General standard laboratory glassware, Dropsens μ Stat200 (Metrohm), and SPCE (Metrohm) were used for preparation and apparatus.

2.2. Modification of Screen-Printed Carbon Electrode

A 5.40 g of FeCl₃.6H₂O and 2.78 g of FeSO₄.7H₂O each were dissolved in 25 mL of distilled water. The two solutions were mixed and added to 100 mL of 1% chitosan solution while stirred at 1000 rpm for 20 minutes at 40 °C. After 20 minutes, the reaction was heated to 60 °C, and then 6 mL of Na-TPP solution was added while continuously stirred at 1000 rpm for 3 hours. 40 mL of 25% NH₄OH was then added dropwise into the solutions. The Fe₃O₄-chitosan was washed 3 times with distillate water and then separated by a magnetic field. The SPCE was coated with Fe₃O₄-chitosan and then heated at 30 °C for 5 minutes to form a thin film.

3. RESULTS AND DISCUSSION

3.1. Screen-Printed Carbon Electrode Modification

The sensor was made by modifying the screen-printed electrode using synthetic Fe_3O_4 -chitosan. Figure 1a shows the SEM image of Fe_3O_4 -chitosan; the particles appear to form aggregates. This is due to its in-situ synthesis. Based on the EDX spectrum (Figure 1-b), it is confirmed that the primary component is iron, which is 70.58%. Thus, Fe_3O_4 is confirmed to be in the Fe_3O_4 -chitosan modifier. Modified SPCE can be evaluated by cyclic voltammetry (CV) compared with SPCE without modification, in which both for the blank solution are a mixture of electrolytes KCl and HCl, each 0.05 M.

Figure 2 presents the voltammogram of blank solutions (a) and (b) 2 ppm Cr(VI) solution. The peak of the CV wave in unmodified SPCE is estimated to be an oxidation of Cl^- to Cl_2 , while in the modified SPCE, it is the oxidation of Fe^{2+} to Fe^{3+} ($E^{\circ}=0.77$ Volt vs SHE). The 2 ppm Cr(VI) solution did not significantly affect the peak potential (Ep) shift, but there was an increase in the peak current (Ip) in the unmodified SPCE. The Ep shift and Ip increase in the modified SPCE is probably due to 2 ppm Cr(VI) (Table 1).

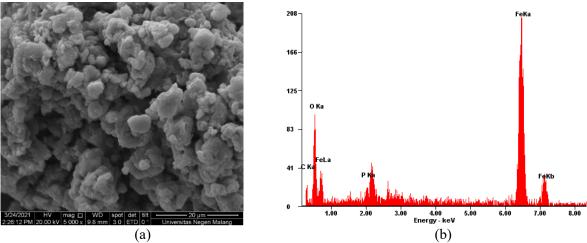


Figure 1. SEM image of Fe₃O₄-Chitosan surface (a); EDX spectrum of the elements in the sample (b).

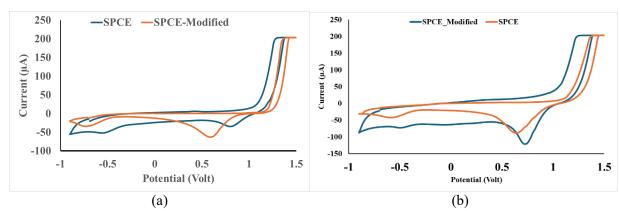


Figure 2. CV voltammogram of 0.05 M KCl-HCl solution (a) 2 ppm of Cr(VI) solution in 0.05 M KCl-HCl (b), in unmodified and modified SPCE, respectively.

TABLE I. Effect of SPCE modification by Fe₃O₄-chitosan on E_p shift and I_p increase in cyclic voltammetry.

Electrode	E_{p} (Volt)			$I_{p}\left(\mu A\right)$			
Electrode	Blank	Cr(VI) 20 ppm	ΔE_p	Blanko	Cr(VI) 20 ppm	ΔI_p	
Unmodified SPCE	0.61	0.63	0.02	52.79	73.40	20.62	
Modified SPCE	0.85	0.74	-0.11	25.43	94.05	68.62	

Hexavalent chromium, in acidic media such as $HCrO_4$, can be reduced to Cr(III) and involves three electrons ($E^\circ = 1.35$ volts vs SHE). In this study, the supporting electrolyte was a mixture of KCl and HCl. Electrolyte concentration greatly influences the peak potential and peak current of the voltammogram. As shown in Figure 3, the peak that appears on the cyclic voltammetry for the blank is the peak of the oxidation of Cl^- to Cl_2 ($E^\circ = 1.36$ volts vs. SHE). There was a shift in peak potential and an increase in current in the 2 ppm Cr(VI) solution compared to the blank, but in 0.05 M KCl-HCl, it was more significant than that of 0.01 M KCl-HCl (Table 2).

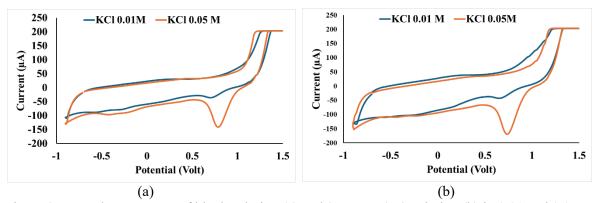


Figure 3. CV voltammogram of blank solution (a) and 2 ppm Cr(VI) solution (b) in 0.01 and 0.05 M KCl-HCl measured by the modified-SPCE.

TABLE II. Effect of electrolyte concentration on E_p shift and I_p increase in the cyclic voltammetry.

	Ер					
[KCl-HCl] (M)	Blank	2 ppm	ΔEp	Blank	2 ppm	ΔIp
0.01	0.76	0.7	-0.06	13.97	22.08	8.10
0.05	0.86	0.77	-0.09	108.81	136.30	27.49

Differential pulse voltammetry (DPV) measurements were also carried out for analysis purposes. The DPV voltammogram is shown in Figure 4 for both the blank and the 2 ppm Cr(VI) solutions, compared between the unmodified and modified SPCE. In Figure 4, the voltammogram of the blank solution in unmodified SPCE has a higher peak current than the modified SPCE. Thus, the presence of Fe_3O_4 -chitosan can decrease the background current (Figure 4a). In Figure 4b, it can be seen that the voltammogram in the modified SPCE is more perfect for Cr(VI) 2 ppm in KCl-HCl 0.05 M. This shows that Fe_3O_4 -chitosan can increase the peak current, kinetics Fe_3O_4 -chitosan in SPCE can accelerate the mass transport of Cr(VI) from the bulk solution to the electrode surface. Chitosan in an acidic environment will be positively charged on its amine group, so it can adsorb Cr(VI), which is negatively charged, so it is estimated that there are two mechanisms of Cr(VI) transport: adsorption and diffusion. But this needs to be proven further. The difference in E_p of blank solution with 2 ppm Cr(VI) solution is higher in unmodified SPCE than in modified SPCE, likewise with each difference in peak current (Table 3).

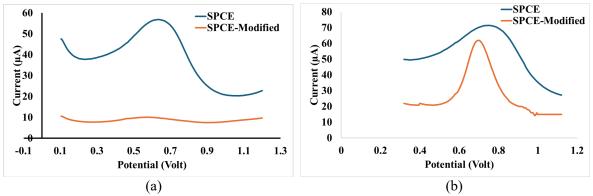


Figure 4. DPV voltammogram of 0.05 M KCl-HCl solution (a) and 2 ppm of Cr(VI) solution in 0.05 M KCl-HCl (b), in unmodified and modified SPCE, respectively.

TABLE III. Effect of SPCE modification by Fe_3O_4 -chitosan on E_p shift and I_p increase in differential pulse voltammetry.

Electrode	E _p (Volt)			I _p (μA)		
Liectiode	Blank	2 ppm	ΔE_p	Blank	2 ppm	ΔI_p
Unmodified SPCE	0.620	0.765	0.145	18.83	31.18	12.35
Modified SPCE	0.626	0.715	0.089	9.50	42.39	32.89

Based on the CV and DPV voltammograms, it is suggested that the voltammetric determination of Cr(VI) can be carried out indirectly using an electrolyte or ion easily oxidized by $Cr_2O_7^{2-}$ in an acidic environment. The results showed that Cl^- was more easily oxidized by the dichromate, but the system was easier to reduce the ions in the working electrode (SPCE). In this case, the E_p is present at 0.7 volts, which is the reduction potential from Fe^{3+} to Fe^{2+} . Therefore, it is most likely that Fe^{3+} is reduced by the system and then re-oxidized by dichromate. Hence, the increase in dichromate concentration is directly proportional to the peak current. It is also noticed that the baseline current of the voltammogram increases proportionally with the dichromate concentration (Figure 5 and Figure 6). Based on EDX measurement,

the modifier contains 70% iron, of which 2/3 of it is Fe³⁺. Due to only a few Fe₃O₄ on the SPCE surface, the concentration of dichromate that can be determined is limited to low concentrations.

3.2. Analytical Parameters

As shown in Figure 4, the better concentration of supporting electrolyte is KCl-HCl 0.05 M, although the background current is considerably high, a normalization can be done. The concentrations of Cr(VI) are varies from 1 to 9 ppm, the sensitivity is 7.3 uA/ppm (Figure 5), which may be due to the limitations of the apparatus. Therefore, to increase the sensitivity, both the Cr(VI) and the electrolyte concentration were reduced by 10 times (Figure 6).

As shown in Figure 6, the sensitivity increases three times in the Cr(VI) concentration range of 0-1 ppm in 0.005 M KCl-HCl supporting electrolyte. The SPCE-Fe₃O₄-Chitosan sensitivity in the Cr(VI) concentration is 21.86 uA/ppm, with a detection limit of 0.2 ppm. This result was obtained in measurement conditions: pulse amplitude is 0.1 volts with an amplitude time of 20 msec, and scan rate is 0.1 mV/sec.

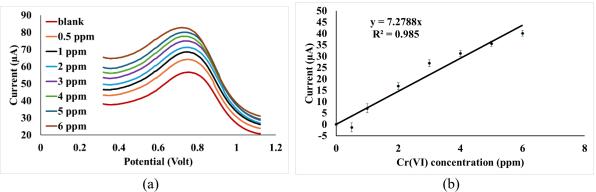


Figure 5. DPV voltammogram of Cr(VI) in 0.05 M KCl-HCl solution (a) and standard curve of Cr(VI) (b), measured by SPCE-Fe₃O₄-Chitosan.

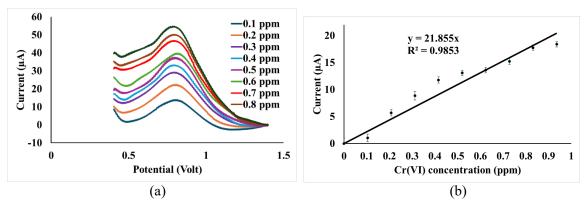


Figure 6. DPV voltammogram of Cr(VI) in 0.005 M KCl-HCl solution (a) and standard curve of Cr(VI) (b), measured by SPCE-Fe₃O₄-Chitosan.

TABLE IV. Concentration of Cr(VI) of the sample, standard addition sample (A) compared with the Cr(VI) concentration of standard plus sample (B).

Sample	Standard	Current (uA)		Concentration of Cr(VI) (ppm)			
	concentration (ppm)		Sample +	Sample -	Sample + Standard		Recovery (%)
			Standard	Sample -	A	В	- (/0)

1	0.8	4.33	19.69	0.20	0.9	1.0	90.28
2	0.8	12.69	27.06	0.58	1.2	1.4	89.68
3	0.8	4.49	22.26	0.21	1.0	1.0	101.29
4	0.8	1.09	16.56	0.05	0.8	0.8	89.17
5	0.8	7.90	23.53	0.36	1.1	1.2	92.71
6	1.2	13.54	37.27	0.62	1.7	1.8	93.73
7	1.2	1.09	25.92	0.05	1.2	1.2	94.90
8	1.2	0.88	26.44	0.04	1.2	1.2	97.56
9	1.2	8.73	33.54	0.40	1.5	1.6	95.95
10	1.2	3.37	28.70	0.15	1.3	1.4	96.98

Validation was carried out on ten aqueous sample solutions and addition standard solutions of Cr(VI) 0.8 and 1.2 ppm, in which each measurement was repeated 5 times. Calculation of the Cr(VI) concentrations was based on the standard curve in Figure 6(b). Using the same standard curve in Figure 6(b), the limit of detection (LoD) of the DPV method can also be calculated for the measurement of Cr(VI) in 0.005 M KCl-HCl. The LoD is calculated based on the assumption of a normal distribution where y = yB + 3SD. The result of the LoD calculation is 0.2 ppm. Table 4 also reported that the recovery or accuracy ranges from (89 - 101) %, averaging 94%.

4. CONCLUSION

Screen-printed carbon electrodes can be modified by Fe₃O₄-chitosan for voltammetric determination of Cr(VI). The Fe₃O₄-chitosan contains 70% iron, which is dominated by Fe(III). Voltammetry modification of SPCE with Fe₃O₄-chitosan can shift the potential and current increase its peak. The average reduction peak potential of the modified SPCE is 0.825 volts in the concentration range of 0.1 – 1.0 ppm Cr(VI) in 0.005 M KCl-HCl solutions. The operating conditions of the instrument were at 0.1 volts pulse amplitude with an amplitude time of 20 msec and a scan rate of 0.1 mV/sec. This method has a sensitivity of 21.86 ppm/μA, a detection limit of 0.2 ppm, and an average accuracy of 94%.

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References

- [1] "Chromium in Drinking-water Background document for development of WHO Guidelines for Drinking-water Quality", pp. 1-6, 2003.
- [2] S. Kristianto, S. Wilujeng, & D. Wahyudiarto, "Analisis Logam Berat Kromium (Cr) Pada Kali Pelayaran Sebagai Bentuk Upaya Penanggulang Pencemaran Lingkungan Di Wilayah Sidoarjo", *Biota*, vol. 3, no 2, pp.66-70, 2017.
- [3] L. L. Wang, J. Q. Wang, Z. X. Zheng, & P. Xiao, "Cloud point extraction combined with high-performance liquid chromatography for speciation of chromium(III) and chromium(VI) in environmental sediment samples", *J Hazard Mater*, vol. 177, no 1–3, pp.114-8, 2010.
- [4] N. Zhang, J. S. Suleiman, M. He, & B. Hu, "Chromium(III)-imprinted silica gel for speciation analysis of chromium in environmental water samples with ICP-MS detection", *Talanta*, vol. 75, no 2, pp. 536-543, 2008.
- [5] S. Yalçin & R. Apak, "Chromium(III, VI) speciation analysis with preconcentration on a maleic acid-functionalized XAD sorbent", em *Analytica Chimica Acta*, vol.505, no. 1, pp.25-35, 2004.
- [6] K. Kiran, K. S. Kumar, B. Prasad, K. Suvardhan, R. B. Lekkala, & K. Janardhanam, "Speciation determination of chromium(III) and (VI) using preconcentration cloud point extraction with flame atomic absorption spectrometry (FAAS)", *J Hazard Mater*, vol. 150, no 3, pp. 582-586, 2008.

- [7] R. E. Shaffer, J. O. Cross, S. L. Rose-Pehrsson, & W. T. Elam, "Speciation of chromium in simulated soil samples using X-ray absorption spectroscopy and multivariate calibration", *Anal Chim Acta*, vol. 442, no 2, pp. 295-304, 2001.
- [8] E. Fabian Ifeanyi & A. Peter Chikezie, "Colorimetric determination of chromium in aluminium alloys by diphenylcarbazide method", *Open Journal of Chemistry*, vol. 5, no 1, pp.9-12, 2019.
- [9] A. Wiryawan, R. Retnowati, P. Burhan, & S. Syekhfani, "Method Of Analysis For Determination Of The Chromium (Cr) Species In Water Samples By Spectrophotometry With Diphenylcarbazide", *Journal of Environmental Engineering and Sustainable Technology*, vol. 5, no 1, pp. 37-46, 2018.
- [10] A. Lace, D. Ryan, M. Bowkett, & J. Cleary, "Chromium monitoring in water by colorimetry using optimised 1,5-diphenylcarbazide method", *Int J Environ Res Public Health*, vol. 16, no 10, pp. 1-15, 2019.
- [11] S. S. M. Hassan, M. S. El-Shahawi, A. M. Othman, & M. A. Mosaad, "A potentiometric rhodamine-B based membrane sensor for the selective determination of chromium ions in wastewater", *Analytical Sciences*, vol. 21, no 6, pp. 673-8, 2005.
- [12] A. Yari & H. Bagheri, "Determination of Cr(VI) with selective sensing of Cr(VI) anions by a PVC-membrane electrode based on quinaldine red", *Journal of the Chinese Chemical Society*, vol. 56, no 2, pp. 289-295, 2009.
- [13] Y. W. Choi & S. H. Moon, "Determination of Cr(VI) using an ion selective eletrode with SLMs containing Aliquat336", *Environ Monit Assess*, vol. 92, no 1–3, pp. 163-78, 2004.
- [14] Y. W. Choi, N. Minoura, & S. H. Moon, "Potentiometric Cr(VI) selective electrode based on novel ionophore-immobilized PVC membranes", *Talanta*, vol. 66, no 5, pp. 1254-1263, 2005.
- [15] D. Li, J. Li, X. Jia, Y. Xia, X. Zhang, & E. Wang, "A novel Au-Ag-Pt three-electrode microchip sensing platform for chromium(VI) determination", *Anal Chim Acta*, vol. 804, no. 1, pp. 98-103, 2013.
- [16] S. A. Tukur, N. A. Yusof, & R. Hajian, "Linear sweep anodic stripping voltammetry: Determination of Chromium (VI) using synthesized gold nanoparticles modified screen-printed electrode", *Journal of Chemical Sciences*, vol. 127, no 6, pp. 1075–1081, 2015.
- [17] D. C. Prabhakaran, J. Riotte, Y. Sivry, & S. Subramanian, "Electroanalytical Detection of Cr(VI) and Cr(III) Ions Using a Novel Microbial Sensor", *Electroanalysis*, vol. 29, no 5, pp. 1222-1231, 2017.
- [18] Y. Liu, G. Gao, J. Hu, & X. Zou, "Electrodeposited AuNPs/rGO nanocomposite as sensor for Cr(VI) determination in water", *Int J Electrochem Sci*, vol. 13, no 12, pp. 11853-11866, 2018.
- [19] C. B. Breslin, D. Branagan, & L. M. Garry, "Electrochemical detection of Cr(VI) with carbon nanotubes decorated with gold nanoparticles", *J Appl Electrochem*, vol. 49, no 2, pp.195-205, 2019.
- [20] W. T. Dinbore, W. C. Dabbo, & A. P. Washe, "Differential pulse voltammetric determination of hexavalent chromium using nickel hexacyanoferrate modified glassy carbon electrode", *Sustainable Environment*, vol. 7, no 1, pp. 1-11, 2021.
- [21] A. A. Nurillah, A. Mulyasuryani, & H. Sulistyarti, "Modification of Glassy Carbon Electrodes on Starch-Based for Detection of Chromium Hexavalent", *Jurnal Kimia Valensi*, vol. 8, no 2, pp. 146-152, 2022.
- [22] P. C. Bandara, J. Peña-Bahamonde, & D. F. Rodrigues, "Redox mechanisms of conversion of Cr(VI) to Cr(III) by graphene oxide-polymer composite", *Sci Rep*, vol. 10, no 1, pp. 1-8, 2020.
- [23] R. Andawiyah, A. Mulyasuryani, & H. Sulistyarti, "Voltammetric Determination of Paracetamol using Polyvinyl Alcohol (PVA)-Fe₃O₄ Modified Glassy Carbon Electrode", *IOP Conference Series: Materials Science and Engineering*, vol. 833, no. 1, pp. 1-8, 2020.
- [24] R. E. K. Billah *et al.*, "A comparative study on hexavalent chromium adsorption onto chitosan and chitosan-based composites", *Polymers (Basel)*, vol. 13, no 19, pp. 1-15, 2021.
- [25] N. Othman, M. J. Masarudin, C. Y. Kuen, N. A. Dasuan, L. C. Abdullah, & S. N. A. M. Jamil, "Synthesis and optimization of chitosan nanoparticles loaded with 1-ascorbic acid and thymoquinone", *Nanomaterials*, vol. 8, no 11, pp. 2-19, 2018.

- [26] D. J. Sullivan, M. Cruz-Romero, T. Collins, E. Cummins, J. P. Kerry, & M. A. Morris, "Synthesis of monodisperse chitosan nanoparticles", *Food Hydrocoll*, vol. 83, no. 10, pp. 355-364, 2018.
- [27] M. Agarwal, M. K. Agarwal, N. Shrivastav, S. Pandey, R. Das, & P. Gaur, "Preparation of Chitosan Nanoparticles and their In-vitro Characterization", *International Journal of Life-Sciences Scientific Research*, vol. 4, no 2, pp. 1713-1720, 2018.